

# **Technical Information**

### **Bismuth Sulphite Agar Modified**

### Product Code: DM 2004

**Application:** - Bismuth Sulphite Agar Modified is recommended for the selective isolation and preliminary identification of *Salmonella* Typhi and other Salmonellae from pathological materials, sewage, water supplies, food etc.

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Ingredients	Gms / Litre
Peptic digest of animal tissue	5.000
Beef extract	5.000
Dextrose	5.000
Disodium phosphate	4.000
Ferrous sulphate	0.300
Bismuth sulphite indicator	8.000
Brilliant green	0.016
Agar	12.700
Final pH ( at 25°C)	7.6±0.2

## **Principle & Interpretation**

The Salmonellae constitute the most taxonomically complex group of bacteria among *Enterobacteriaceae* <sup>(1)</sup>. Human *Salmonella* infections are most commonly caused by ingestion of food, water or milk contaminated by human or animal excreta. Humans are the only reservoirs of *S.* Typhi <sup>(2)</sup>. Four clinical types of *Salmonella* infections have been described namely gastroenteritis, bacteremia or septicemia, enteric fever and a carrier state <sup>(3)</sup>. Of the various media employed for the isolation and preliminary identification of Salmonellae, particularly *Salmonella* Typhi; Bismuth Sulphite Agar is the most suitable one <sup>(4)</sup>.

Bismuth Sulphite Agar, Modified is a modification of the original formulation of Wilson and Blair Medium <sup>(5)</sup>. It is also recommended for the isolation of *Salmonella* Typhi and other Salmonella <sup>(6, 7)</sup>.

S. Typhi, S. Enteritidis and S. Typhimurium typically grow as black colonies with a surrounding metallic sheen resulting from hydrogen sulphide production and reduction of sulphite to black ferric sulphide. Salmonella Paratyphi A grows as light green colonies. Bismuth Sulphite Agar may be inhibitory to some strains of Salmonella like S. Sendai, S. Berta S. Gallinarum, S. Abortus-equi and therefore should not be used as the sole selective medium for these organisms. Shigella species are mostly inhibited on this medium; exceptions being S. flexneri and S. sonnei (8). Also this medium favors use of larger inoculums as compared to other selective media, as it has unique inhibitory action toward gram-positive organisms and coliforms.

Peptic digest of animal tissue and beef extract serve as sources as carbon, nitrogen, vitamins and essential growth factors. Dextrose is the carbon source. Disodium phosphate maintains the osmotic equilibrium. Bismuth sulphite indicator along with brilliant green inhibits the intestinal gram-positive and gram-negative bacteria. Ferrous sulphate aids in detection of hydrogen sulphide production. Clinical samples can be directly used to inoculate Bismuth Sulphite Agar. In case of food samples, pre enrichment of the sample is done prior to inoculation.

### Methodology

Suspend 40 grams of powder media in 1000 ml distilled water. Shake well & heat to boiling to dissolve the medium completely. DO NOT
STERILIZE IN AUTOCLAVE or by fractional sterilization since overheating may destroy the selectivity of the medium.

The sensitivity of the medium depends largely upon uniform dispersion of precipitated bismuth sulphite in the final gel, which should be dispersed before pouring into the sterile Petri plates.





### **Quality Control**

#### Physical Appearance

Light yellow to greenish yellow homogeneous free flowing powder

#### Gelling

Firm, comparable with 1.27% Agar gel.

#### Colour and Clarity of prepared medium

Greenish yellow coloured opalescent with flocculent precipitate forms in Petri plates.

#### Reaction

Reaction of 4.0% w/v aqueous solution at 25°C. pH: 7.6±0.2

#### pH range

7.40-7.80

#### CulturalResponse/Characteristics

DM2004: Cultural characteristics observed after an incubation at 35-37°C for 40-48 hours.

	Organism	Inoculum (CFU)	Growth	Recovery	Colour of Colony
Enterobacter aerogenes ATCC 13048		50-100	None-poor	<=10%	browngreen(depends on inoculum density)
	Enterococcus faecalis ATCC 29212	>=10 <sup>3</sup>	Inhibited	0%	,
	Escherichia coli ATCC 25922	50-100	None-poor	<=10%	browngreen(depends on inoculum density)
	Salmonella Typhi ATCC 19430	50-100	Good-luxuriant	>=50%	black with metallic sheen
	Salmonella Paratyphi B ATCC 8759	50-100	Good-luxuriant	>=50%	black with metallic sheen
	Salmonella Enteritidis ATCC 13076	50-100	Good-luxuriant	>=50%	black with metallic sheen
	Shigella flexneri ATCC 12022	50-100	None-poor	<=10%	Brown
	Salmonella Typhimurium ATCC 14028	50-100	Good-luxuriant	<=50%	Black with metallic sheen

## Storage and Shelf Life

**Dried Media:** Store below 30°C in tightly closed container and use before expiry date as mentioned on the label.

Prepared Media: 2-8° in sealable plastic bags for 2-5 days

## **Further Reading**

- 1. Tindall B. J., Crimont P. A. D., Gorrity G. M., EUZESY B. P., 2005, Int. J. Sys. Evol. Microbiol., 55:521
- 2. Koneman E. W., Allen S. D., Janda W. M., Schreckenberger P. C., Winn W. C. Jr., 1992, Colour Atlas and Textbook of Diagnostic Microbiology, 4 th Ed., J. B. Lippinccott Company
- 3. Mandell G. L., Douglas R. G. Jr., Bennet J. E., (Eds.), 1985, Principles and Practice of Infectious Diseases, 2nd Ed., 660-669, John Wiley & Sons New York.
- 4. Gunter and Tuft, 1939, J. Lab. Clin. Med., 24:461.
- 5.Wilson and Blair, 1927, J. Hyg., 26:374
- 6. Anon, 1981, Int. Standard ISO 6579-198 1, Geneva. International Organization for Standardization.
- 7. ICMSF, 1978, Microorganisms in Food, 2nd Edi, University of Toronto Press, Ontario.
- 8. MacFaddin J. F., 2000, (Ed.), Biochemical Tests for Identification of Medical Bacteria, 3rd Edition, Lippincott, Williamss & Wilkins, New York.

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